

Evaluation of toxicological effects of POPs using primary fish cell cultures

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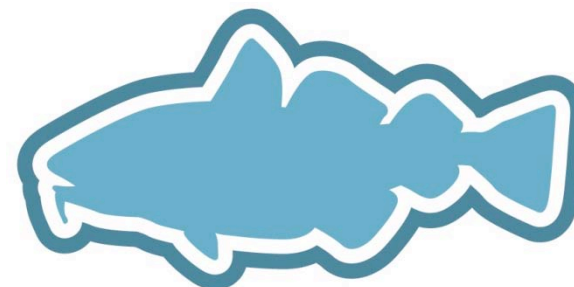
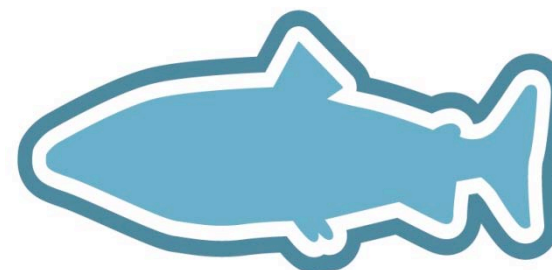
National Institute of Nutrition and Seafood Research





Primary fish cells

- 90% of all experimental animals used in research in Norway are fish
- Important to develop *in vitro* fish bioassays
 - refine and replace existing fish experimental protocols
 - reduce the number of fish used in research
- It is important to develop species specific *in vitro* systems
 - especially important for farmed fish species with commercial interest





The nutritional composition of the media could have an effect on toxicological responses

- **Supplementation of serum vs. serum-free media**

- Serum

- Content of nutrients, growth factors, hormones, minerals and lipids are of importance for cell maintenance
 - Concentration and action of the constituents of serum are not fully determined
 - Problem with standardization of experiments and between laboratories due to batch effects and accessibility of different serum
 - Use of high serum concentration can reduce chemical bioavailability and toxic responses. Can have an effect on downstream processing

- Serum-free media

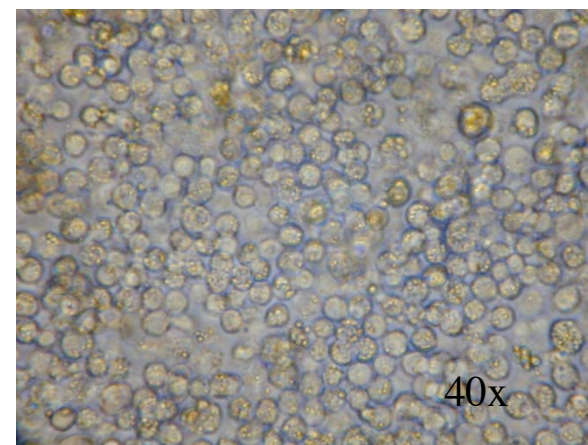
- More control on proliferation and differentiation, and growth optimization for specific cell types
 - Reduced cell growth and selection of resilient cells

- **Fish cultures:**

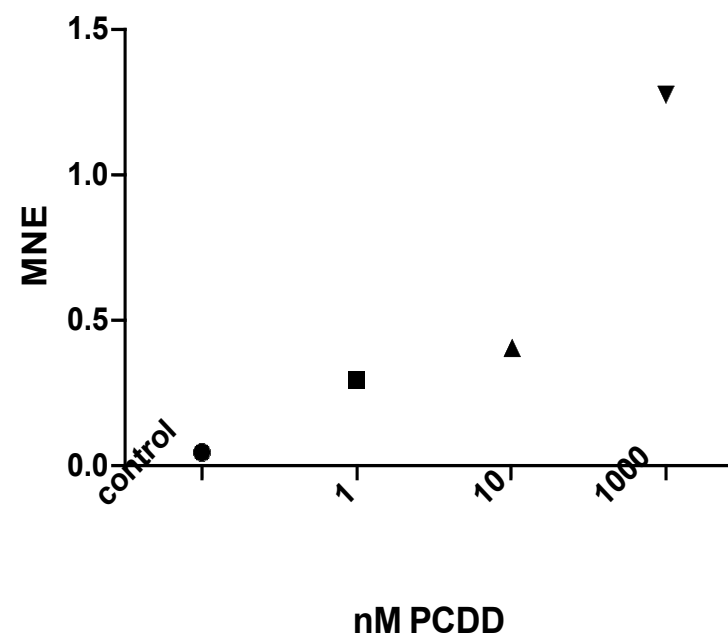
- FBS (fetal bovine serum) is frequently used
 - Use of fish serum might be a better alternative
 - Need for standardization of experiments and between laboratories

Primary Atlantic cod hepatocyte cultures

- Important to develop primary hepatocyte cultures since liver is the main biotransformation organ
- It has been problematic to establish cod hepatocyte cultures due to the high fat content in cod liver
- Intact cod hepatocytes have been isolated from mature individuals
 - 1,2,3,7,8'-PeCDD exposed cod cells induced CYP1A



CYP1A





**Factorial design applied for multiple-endpoint
toxicity evaluation in Atlantic salmon
(*Salmo salar* L.) hepatocytes**

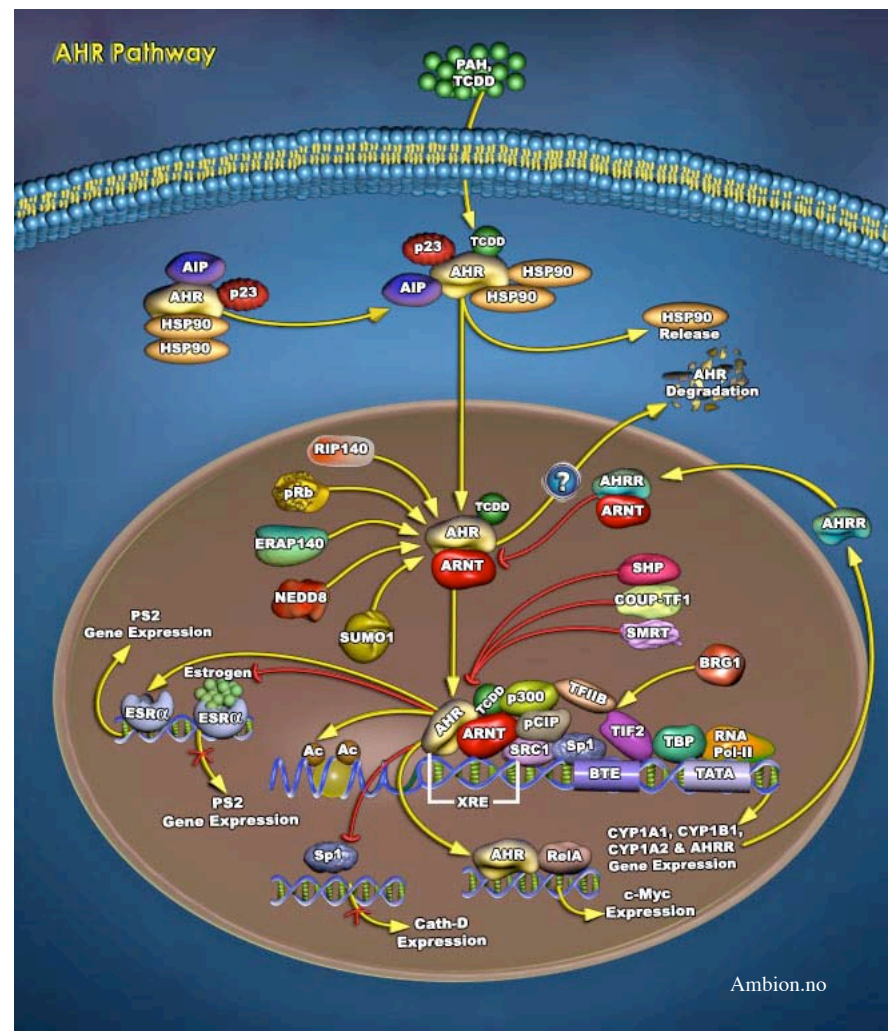
The organic pollutants Polychlorinated dibenzo-*p*-dioxins (PCDDs), Polychlorinated dibenzofurans (PCDFs) and Polychlorinated biphenyls (PCBs)

- **PCDD/Fs and PCBs**

- They are still of great concern regards to food safety
 - Relative high concentrations have been measured in farmed fish

- **Mechanism of toxicity**

- **Dioxins and dioxin-like PCBs**
 - Toxicity are induced through the aryl hydrocarbon receptor (AhR)
 - Binding of ligand to AhR results in transcriptional up-regulation of CYP1A and the AhR gene battery (e.g. UDPGT = UGT)
- **Non-dioxin-like PCBs**
 - Toxic responses are induced through other mechanisms



The TEQ method can underestimate toxicity

- **The toxic equivalent (TEQ) method:**

- Interaction between chemicals in mixtures are detected by comparing the theoretical TEQ and experimental TEQ

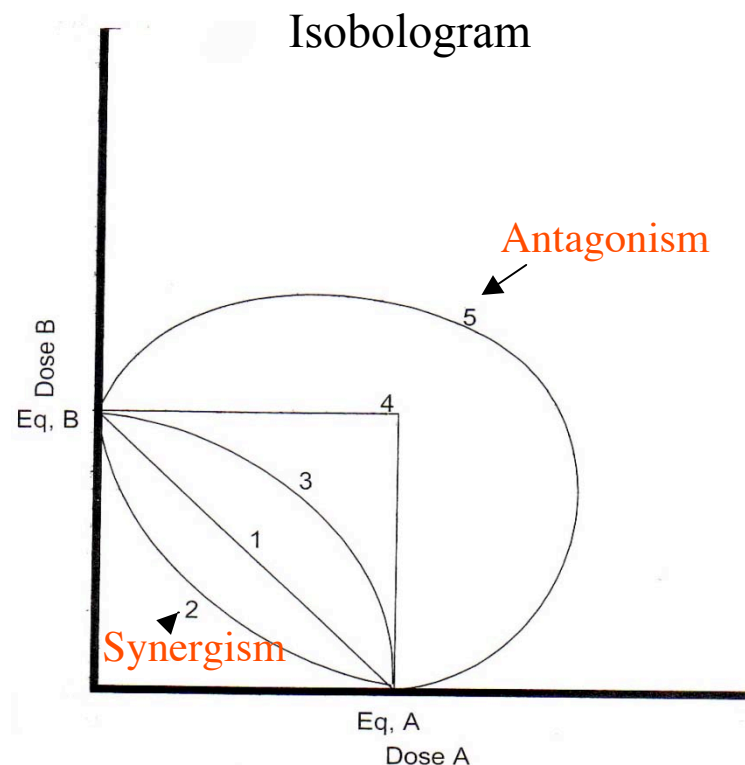
- Assumptions:

- Identical dose-response curves
- dose additivity



Usually not achieved in complex mixtures

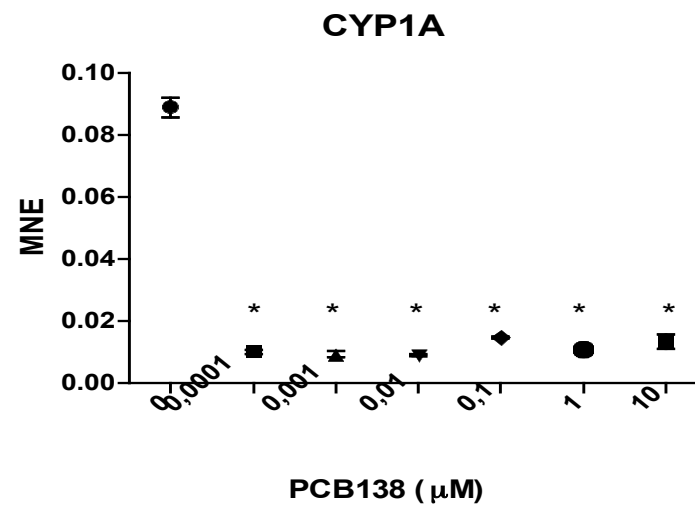
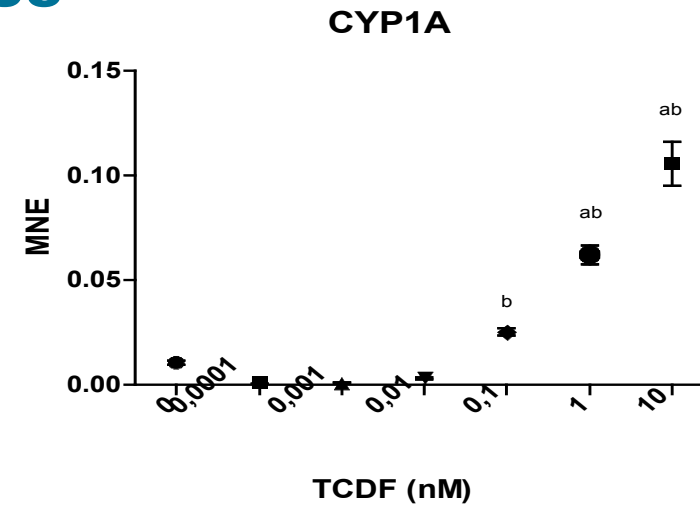
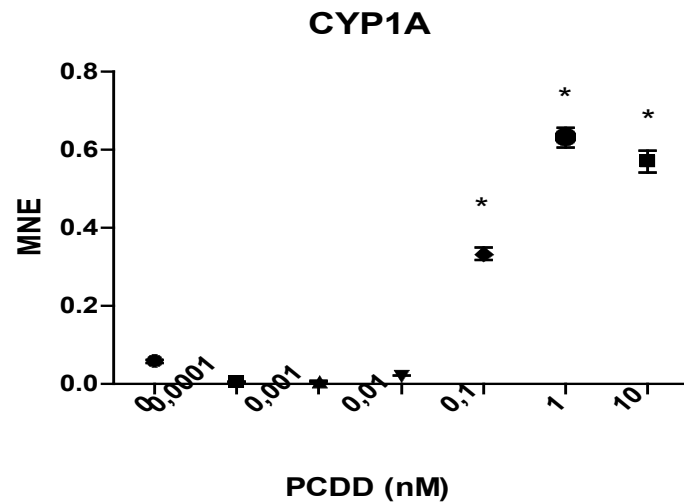
- Essential to develop a alternative method to evaluate occurrence of combine effects between chemicals in mixtures



Combined effects between chemicals in mixtures (Cassee et al., 1999):

- Additivity: Dose additivity (1) and response additivity (3)
- Interaction: Synergism (2) and antagonism (5)

CYP1A dose-response curves in hepatocytes exposed to 1,2,3,7,8-PeCDD, 2,3,7,8-TCDF and PCB138



Factorial design

Ex.no.	PCDD (nM)	TCDF (nM)	PCB138 (μM)
1	0.03	0.1	0.05
2	0.1	0.1	0.05
3	0.03	1	0.05
4	0.1	1	0.05
5	0.03	0.1	1
6	0.1	0.1	1
7	0.03	1	1
8	0.1	1	1
9	0.065	0.55	0.525
10	0.065	0.55	0.525
11	0	0	0

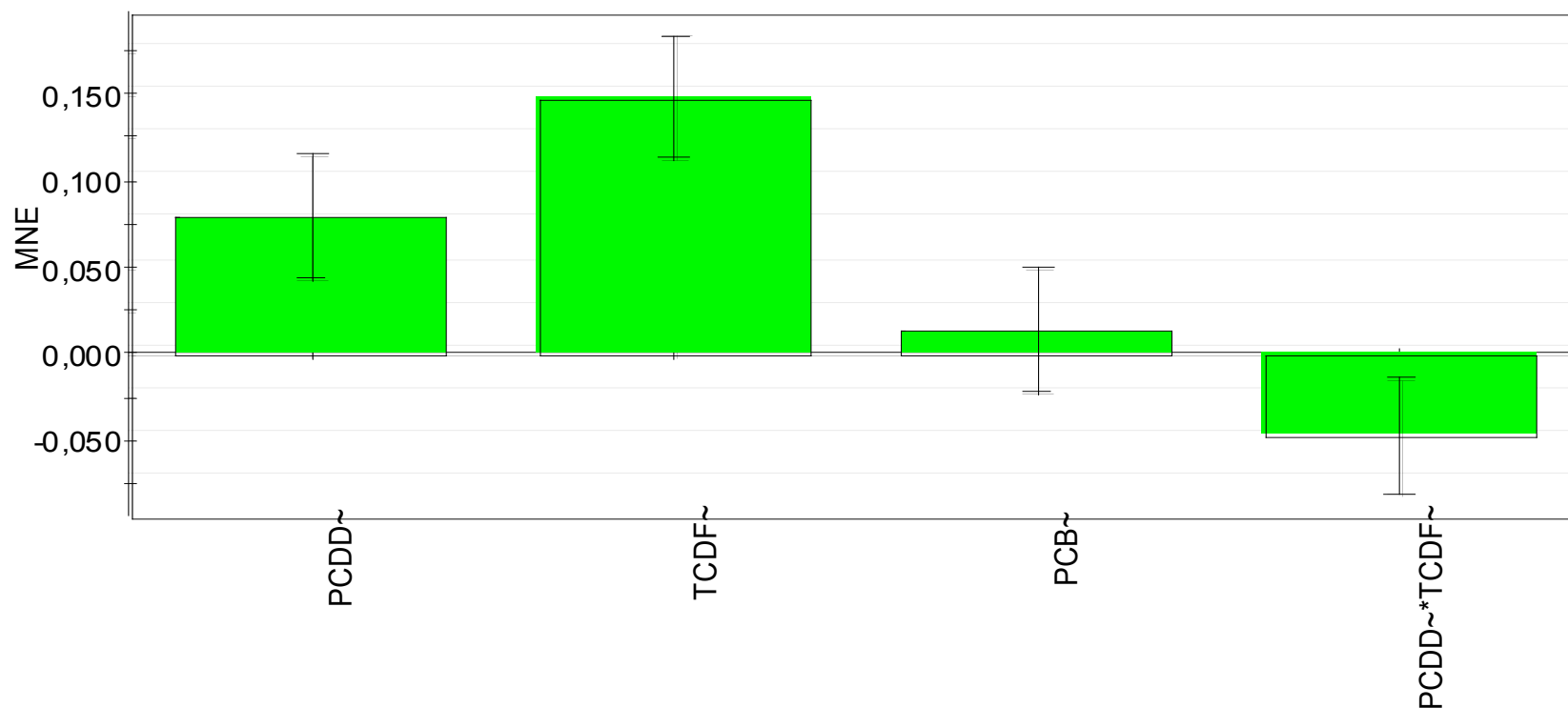
- High
- Middle, center points
- Low
- DMSO control

qPCR:

- Phase 1 and II enzymes: CYP1A, UDPGT and GST
- Cellular stress: HSP70, GR, GPX and Mn SOD
- Apoptosis: p53
- Control genes: Beta-actin, EL and ARP

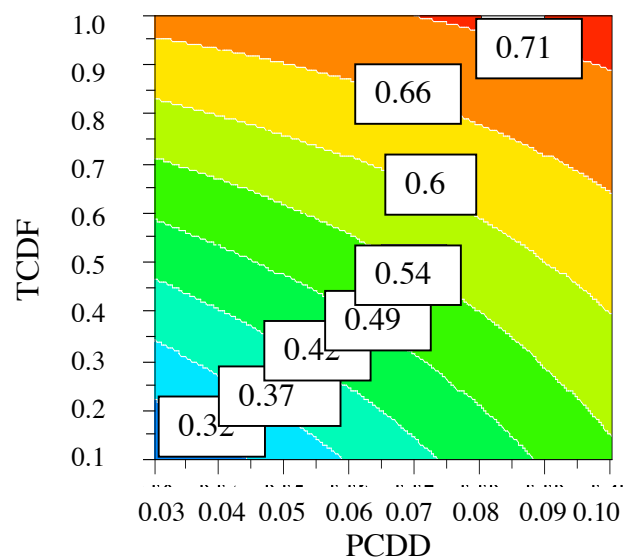
PLS analysis of CYP1A gave a good model with good prediction capability

Scaled & Centered Coefficients for CYP1A~

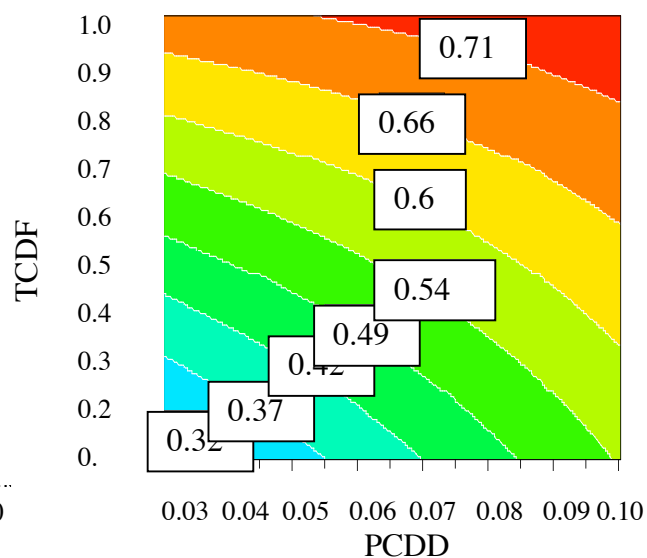


$R^2 = 0.97$ $Q^2 = 0.768$ Conf. lev. = 0.95 1 Comp.

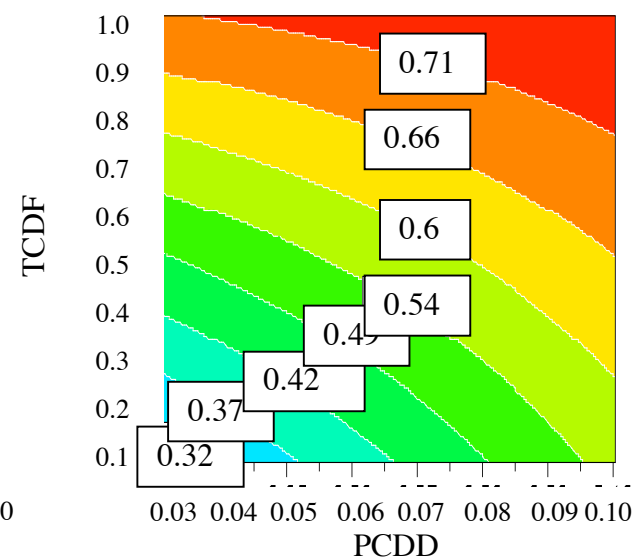
CYP1A contour plot: Antagonism at high concentrations



PCB 138~ 0.05



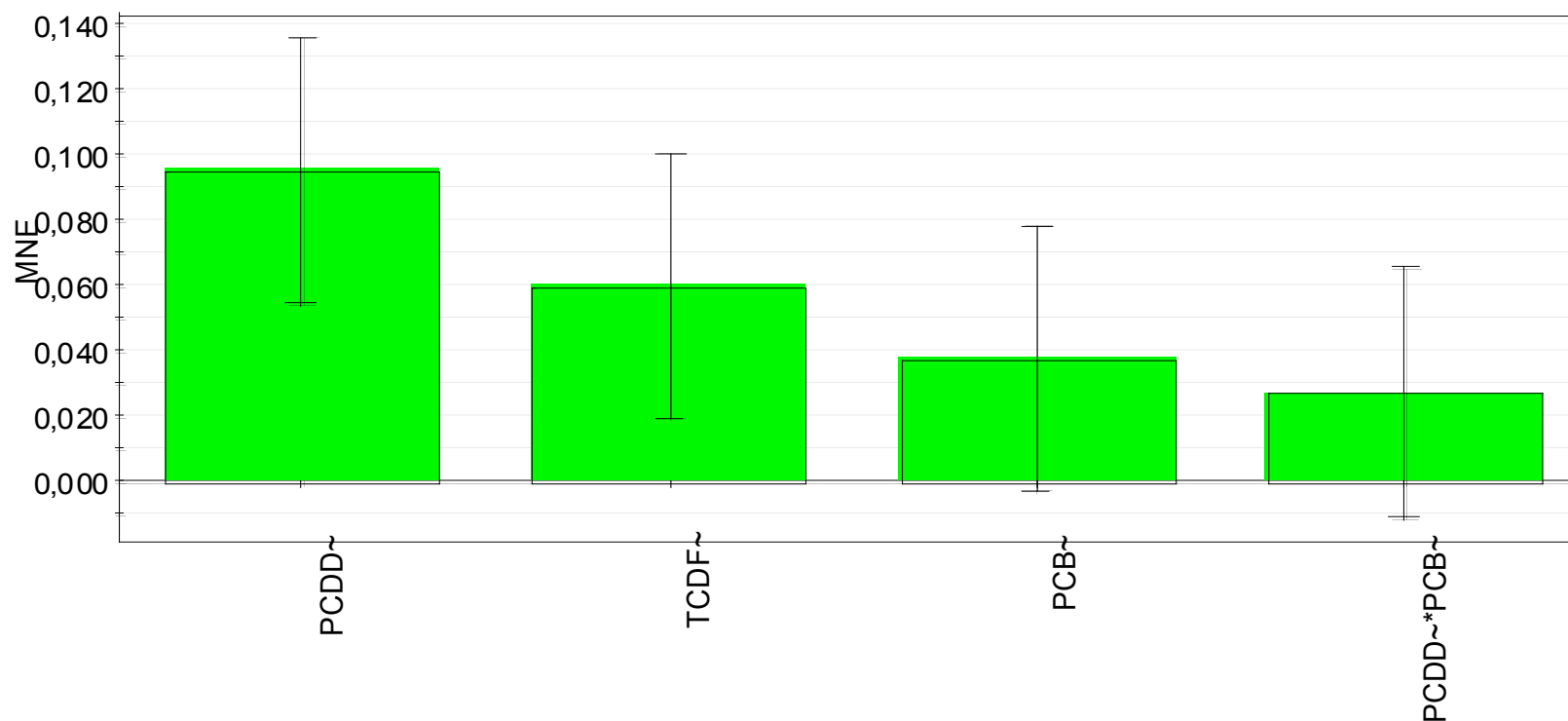
PCB 138~ 0.525



PCB 138~ 1

PLS analysis of UDPGT gave a good model with good prediction capability

Scaled & Centered Coefficients for UDPGT~

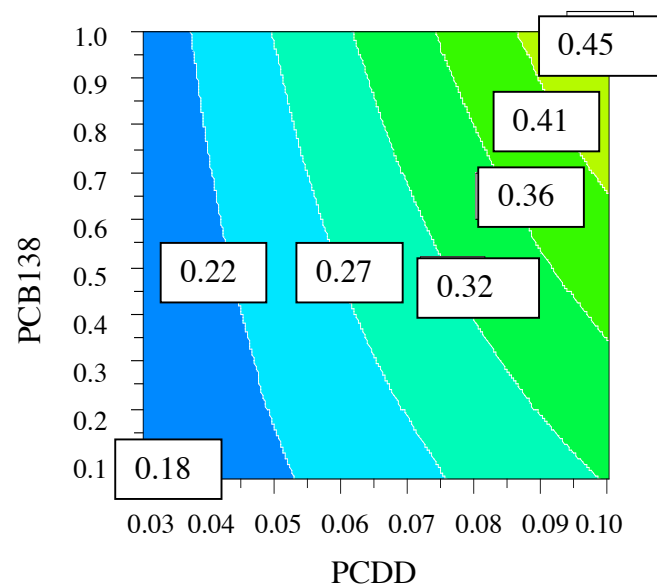


$R^2 = 0.922$

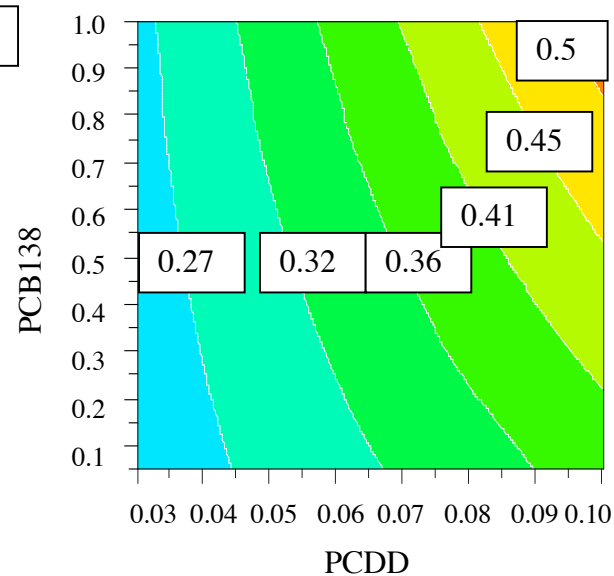
$Q^2 = 0.700$

Conf. lev. = 0.95 1 Comp.

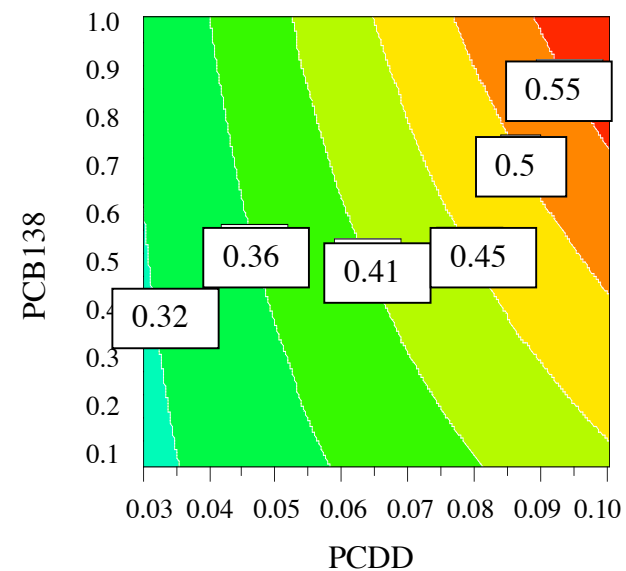
UDPGT contour plot: Synergism at low concentrations



TCDD ~ 0.1



TCDD ~ 0.55



TCDD ~ 1

Synergism and antagonism

- PCB 138 can contribute to the measured CYP1A and UDPGT responses
- The synergistic interaction between PCB 138 and PCDD on UDPGT might counteract the CYP1A antagonistic effect of TCDF and PCDD
 - Possible mechanisms for the synergism:
 - Hepatic retention of potent CYP1A inducers (de Jongh et al., 1993)
 - Higher levels of AhR in the cells (de Jongh et al., 1993; Cherng et al., 2001)
 - And/or higher activation of AhR to a DNA binding form (Cherng et al., 2001)
- Previous studies have detected antagonistic interaction between chemicals on CYP1A at high concentration (Vamvakas et al., 1996; Østby & Krøkje, 2002)
 - Possible mechanisms antagonism:
 - Competitive inhibition (Gooch et al., 1989; Fent & Bättscher, 2000)

Conclusions

- Important to develop *in vitro* fish bioassays
- Intact Atlantic cod hepatocytes have been isolated and PCDD exposed cells induce CYP1A
- Only CYP1A and UDPGT were proved to be good biomarkers for PCDD, TCDF and PCB138
- Using primary fish cell cultures, multivariate data analysis of qPCR data are shown to be a useful tool in toxicological studies
- A multi-endpoint strategy can enhance the quality of risk evaluation of chemical compounds
- But: TEQ is a useful concept in risk assessment of mixtures with dioxin and dioxin-like compounds, until a better evaluation strategy is developed

Thank you for your
attention!

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